



Genotype by Environment Interaction Effects on the Crop of Sugar Beet (*Beta vulgaris* L.) Using Multivariate Analysis

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Evaluation of genotypes under Egyptian desert conditions comes in the first order for the Plant Breeding and Conservation Program of the Desert Research Center (DRC). The objective of this study was to analyze the effect of the genotype by environment interaction of sugar beet across various locations using multivariate models. Data for studied traits of sugar beet were obtained from experiments at three regions: Saint Catherine, South Sinai Governorate (E1); Baloza station, North Sinai Governorate (E2); and East El- Qantra station, El-Ismailia Governorate (E3) in Egypt. All examined traits were significantly impacted ($p < 0.05$ or 0.01) by environment (E), genotypes (G), and their interaction (GEI) using the AMMI model, with the exception of root length/plant by the environments as well as leaves weight/plant and total soluble solids percentage % traits by the genotypes. GEI was partitioned into two principal components (PCs), which were significant for all studied traits ($P < 0.05$ and $P < 0.01$). The highest variability from the total variance was recorded by environmental influences for leaves weight/plant and total soluble solids percentage % traits, as

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well as by genotype effects for the other studied traits. The environmental index showed that some environments were favorable and some environments were unfavorable for the two traits. The highest root weight/plant and most studied traits were noticed in the E2 environment. Based on the GGE model for root weight/plant, the test environments E1 and E2 are more representative and have the greatest ability to discriminate genotypes, thus favoring the selection of superior genotypes. The genotypes G2, G5, and G6 perform best in the E1 and E2 environments as well as are the most productive and stable compared with the other genotypes. According to PCA and cluster analysis, the genotypes G5 and G6 showed the best performance in response to environments and positive association with root weight/plant and most studied traits. Based on the results of statistical methods used in this study, G5 and G6 genotypes should be used in future sugar beet breeding in an effort to improve productivity and sustainable production of sugar beet in Egypt.

Keywords: Sugar beet; ANOVA; GEI; environmental index; multivariate analysis.

1. INTRODUCTION

Sugar beet (*Beta vulgaris* L.) is considered one of the most important sugar crops in many countries all over the world and Egypt. It is the second sugar crop in the world after sugar cane [1,2]. It ranks the first important sugar crop in Egypt and many countries all over the world [3]. Beet growing is becoming more and more important in sugar-producing, the secondary production, fodder and organic matter for the soil [3] as well as in the production of biofuels [4]. In the world, the total area harvested, yield and production of sugar beet were 4439073 ha, 569869 hg ha⁻¹ and 252968843 tons in 2020, respectively. While the total area harvested, yield and production of sugar beet were 263543 ha, 494933 hg ha⁻¹ and 13043612 tons in Egypt [5].

Locations, growth seasons, years, drought conditions, rainfall, the amount of precipitation received throughout each season, temperature, and other environmental factors (non-genetic factors) may have favorable or unfavorable effects on genotypes [6]. According to Falconer and Mackay [7], the phenotypic expression of an individual is determined by both genotype and environmental effects such as locations and/or years, and the phenotypes can be observed, quantified, categorized, or counted. The term "genotype-environment interaction (GEI)" refers to the relationship between a genotype's phenotypic expression and the environment. Breeders assess genotypes in various environments (locations and/or years) to account for GEI effects in order to select those with high and stable performance and higher adaptation, taking GEI effects into account [8]. Stable genotypes were identified with relatively high yield across environments [9], and with insignificant GEI [10]. With respect to sugar beet, the assessment of GEI would be highly beneficial

in identifying traits promoting a better variety of performance in specific environments [11]. Various statistical methods such as parametric, nonparametric statistics, and multivariate models are used for the investigation and interpretation of GEI and evaluation of different genotypes [12]. Among these methods, the additive main effect and multiplicative interaction (AMMI) model and genotype (G) main effect plus genotype-by-environment (GE) interaction (GGE) biplot.

The additive main effect and multiplicative interaction (AMMI) model established by Zobel et al. [13] is the most well-known of the multivariate methods used for the interpretation of GEI data. The AMMI model characterizes genotype and environment main effects using analysis of variance as an additive model, and their interactions using principal components analysis as a multiplicative model (IPCA). The visual method GGE biplot (Genotype and Genotype-Environment Interaction) represents a modification of the AMMI model and gives an extensive explanation of GEI effect [8]. The method is used for visual analysis of multi-environment trial data and it is based on two important concepts: the biplot concept [14] and GGE concept [8]. This method uses a biplot to describe two factors, genotype, and interaction between genotype and environment (G and GEI), thus the name GGE, which can be very useful in the evaluation of the performance of genotypes in different environments [15]. Sugar beet breeders all over the world have been using AMMI analysis of variance and the GGE model to investigate GEI in environmental trials [16-20].

To visualize the outcomes of sugar beet trials, principal component analysis (PCA) and cluster analysis are required, where many researchers have used the PCA to assess the relationship

and diversity between several sugar beet genotypes, in addition to knowing the relationships between yield, its components, and other traits, such as Škrbić et al. [21], Hu et al. [22], Alami et al. [23], Islam et al. [24], Kleuker and Hoffmann [25] and Majumdar et al. [26]. The aim of this work was to study the effect of environment on genotypes of sugar beet under various regions in Egypt, and evaluate the magnitude of genotype x environment interaction using the multivariate methods, thus discovering the most stable sugar beet genotype and determining associations for the studied traits in ten sugar beet genotypes under three different environments.

2. MATERIALS AND METHODS

2.1 Plant Material and Field Trials

The three field experiments were conducted 1) at a local community farm, Saint Catherine (E1), South Sinai, Governorate; 2) at Baloza station (E2), DRC, North Sinai Governorate; and 3) at East El-Qantra station (E3) of DRC, El-Ismailia Governorate in Egypt. Ten sugar beet genotypes were used to evaluate under three locations (environments) during the 2021/2022 growing season. The genotypes used in this study are SK58-4-S4 (G1), SK48-C (G2), SK70 (G3), SK90 (G4), SK44-1-4R2 (G5), SK27-38 (G6), SK27-32 (G7), SK Fc723 (G8), SK44-1-9C1 (G9) and SK17-2-7W (G10). Healthy seeds of sugar beet genotypes were obtained from the Plant Breeding and Conservation Program of Desert Research Center (DRC), Egypt. Sugar beet genotypes were evaluated in Randomized Complete Block Design (RCBD) with three replicates across the three environments. Each replicate included ten plots of genotypes. Each experimental plot had five rows, and the genotypes were planted using standard agronomic practices and proper plant geometry with a row length of 5 m and comprised 25 hills. Under each plot, the row x row and plant x plant distances were 50 and 20 cm, respectively. The crop was sown in one day, and all the recommended cultural practices of sugar beet production in the area were done as needed, under uniform field conditions to minimize environmental variations to the maximum possible extent.

In Table 1, the meteorological data for the three environments is shown as monthly averages for precipitation (mm), average temperature (°C), and relative humidity (%) during the experimental

period (from October to May) during the 2021/2022 growing season. Under the three studied environments, the highest temperature typically occurred in October and May months, while the lowest temperature was recorded in January month. The maximum monthly precipitation rates were recorded at Saint Catherine and Baloza Stations in January, and at East El-Qantra Station in February. The highest values of relative humidity were observed in October, January, and November months at Baloza Station, Saint Catherine, and East El-Qantra Station, respectively. Generally based on the grand mean, Saint Catherine had the lowest average temperature during the study period, followed by Baloza and East El-Qantra Stations. While the greatest values of precipitation and relative humidity were observed in Baloza Station compared with Saint Catherine, and East El-Qantra Station.

2.2 Data Recording

The data on an individual plant basis of the ten genotypes were recorded. Ten guarded plants randomly were collected from each genotype from each replication to evaluate the following traits: leaves weight/plant (g; LW/P), root length/plant (cm; RL/P), root diameter/plant (cm; RD/P), root weight/plant (g; RW/P) and total soluble solids percentage (%; TSS). TSS% was determined by using Hand Refractometer and expressed as a percentage of the juice.

2.3 Statistical Analysis

The analysis of variance by the AMMI model was performed to determine the main and interaction effects of environments and genotypes on evaluated traits. The least significant differences (LSD) at 0.05 and 0.01 levels of probability were also used for means separation and comparison after significance [27]. After determining the significance of the GEI, adaptation ability and phenotypic stability analyses for genotypes studied were performed graphically using the GGE-biplot model [8]. The AMMI model, GGE-biplot, cluster analysis and PCA were done using computer software programs PBSTAT, PAST version 4.03 and OriginPro 2018 version b9.5.0.193.

3. RESULTS AND DISCUSSION

3.1 AMMI Analysis of Variance

Table 2 displays the results of the AMMI analysis of variance of the environments, genotypes, and GEI effects on investigated traits in sugar beet.

The mean squares due to environments, genotypes, and GEI were significant at 0.05 or 0.01 probability levels for all investigated traits, with the exception of RL/P trait by environments, as well as LW/P and TSS % traits by genotypes. A considerable part of the overall variation was formed by the environments for LW/P and TSS% traits, as well as by the genotypes for other studied traits. Curcic et al. [17], Hassani et al. [18] and Bocianowski et al. [20] have previously revealed significant variation between genotypes, environments, and their interactions for all considered traits of sugar beet using AMMI model. Significant variations in the response of genotypes to the effect of environments show the right choice of experimental sites for GEI assessment [18].

After removing sums of squares (SS%) due to error and replication, the greatest contribution to the SS% of the total variance was due to genotypes for root traits including RL/P, RD/P and RW/P, followed by GEI. While, the environments source had recorded the highest contribution to the SS% of the total variance for LW/P and TSS% traits, followed by GEI for LW/P, and genotypes for TSS% in that order. These results indicated that more than 50% of the total variance observed for LW/P and TSS % traits were due to environmental influences, and for RL/P and RD/P traits it was due to the genotype effects. Curcic et al. [16] reported that the environment had the greatest effect on the root yield of sugar beet. On the other hand, the variance due to GEI ranged from 12% for TSS % to 35 for RD/P. The production of sugar beet crop is based on root and its quantitative traits estimated, which is the main cause of the low degree of interaction in this crop [16]. Contrary to other field crops, the sugar beet root grows throughout the vegetative stage of development and does not pass through delicate phases like bolting, flowering, pollination, and seed filling [11]. According to Basford and Cooper [28], the breeders may be able to create more stable genotypes if they have a better grasp of the relative contributions of genotypes, environments, and GEI as sources of variation.

Differences in the variances of the phenotypes produced by the various genotypes can be used to detect the GEI [29]. According to the AMMI analysis, GEI was partitioned into two principal components (PC1 and PC2). The PC1 and PC2 showed significant ($P < 0.05$ or $P < 0.01$) for all examined traits. More than 59% of the total SS from GEI were explained by the PC1 for all

studied traits. In contrast, PC2 contribution ranged from 28% for the LW/P trait to 40% for the RW/P trait. The fact that there was a significant GEI for yield shows that some genotypes were stable while others were unstable [30]. The values of coefficients of variation (CV%) ranged from 3.84% for TSS% trait to 12.88% for LW/P trait. The values of CV% indicate that the genotypes had exploitable genetic variability during selection, also the low CV% proved the accuracy of the sugar beet experiments in three studied environments. Our results of CV% are in accordance with earlier findings on sugar beet crop by Bayomi et al. [31].

3.2 Main Effects of Environments, Genotypes, and Their Interaction on Sugar Beet Traits

Table 3 includes the mean performances of the main effects of environments, genotypes, and their interaction on studied traits as well as the environmental indicator (EI) and coefficient of variation (CV%). The average comparisons of the analyzed traits showed that the evaluated genotypes in each environment differed significantly from one another. Based on the grand mean, the E2 location had the highest mean performances for all studied traits with compared to the other environments, except RL/P trait. The highest mean performances of genotypes over three environments were observed by G9 genotype for LW/P (116.52 g), by G2 genotype for RL/P (16.29 cm) and TSS% (20.11%), and by G5 genotype for RD/P (10.00 cm) and RW/P (886.49 g). With respect to GE interaction, the highest values of mean performance were recorded by the G9 and G4 genotypes for LW/P (163.50 g) and TSS% (23.33%), respectively in E2, as well as by the G2 genotype for RL/P (17.47 cm) and by G5 genotype for RD/P (12.23 cm) and RW/P (1078.67 g) in E2. The differential yield ranking of genotypes across environments is proof that the GEI effect was of the crossover type [32]. According to Thillainathan & Fernandez [33], consistent performance across many environments (locations and/or years) may be the cause of yield stability. LW/P and RW/P traits showed a high CV% in E2 and moderate CV% in E3 and E1, respectively. Opposite, the other studied traits had low CV% values during the studied environments. The estimates of CV% show that the genotypes exhibited genetic variability that may be used during the selection of sugar beet yield under various environmental conditions.

Table 1. Monthly climatic data for the studied environments during 2020/2022 sugar beet growing season

Environments	Climatic data	October	November	December	January	February	March	April	May
Saint Catherine (E1)	Temperature (°C)	18.9	13.4	9.2	7.4	9.4	12.9	17.1	21.1
	Precipitation (mm)	8.0	4.0	3.0	8.0	6.0	6.0	4.0	4.0
	Relative Humidity %	34.0	37.0	38.0	39.0	32.0	27.0	22.0	21.0
Baloza Station (E2)	Temperature (°C)	22.4	18.0	13.7	11.9	13.0	15.6	18.5	21.8
	Precipitation (mm)	7.7	11.7	19.0	26.0	21.0	13.3	5.0	2.3
	Relative Humidity %	68.0	63.7	64.7	66.0	62.7	58.3	55.7	57.3
East El-Qantra Station (E3)	Temperature (°C)	23.5	19.1	15.0	13.3	14.5	17.3	20.5	24.2
	Precipitation (mm)	2.0	2.7	3.3	6.0	6.3	4.0	2.0	0.7
	Relative Humidity %	58.0	58.3	58.0	57.0	52.0	48.3	44.3	43.7

Source: <https://en.climate-data.org>

Table 2. Combined ANOVA with AMMI analysis for studied traits of sugar beet genotypes evaluated across different environments

S.O.V	D.F.	LW/P	RL/P	RD/P	RW/P	TSS %
Environment (E)	2	19584.43**	0.17 ^{NS}	1.25*	95721.29**	144.63**
Replication/E	6	31.89 ^{NS}	0.34 ^{NS}	0.12 ^{NS}	2914.65 ^{NS}	0.66 ^{NS}
Genotype (G)	9	1118.38 ^{NS}	17.60**	8.06*	137451.45*	6.05 ^{NS}
G X E	18	582.60**	2.78**	2.80**	40296.15**	2.92**
PC1	10	752.02**	3.11**	3.09**	42828.81**	3.65**
PC2	8	370.83*	2.36*	2.43**	37130.33**	2.00**
Residuals	54	149.12	1.06	0.30	7373.78	0.51
Contribution to the sums of squares % of total variance explained						
% due to E		57.63	0.13	1.75	7.45	67.65
% due to G		14.81	59.11	51.04	48.14	12.74
% due to G X E		15.43	18.66	35.40	28.23	12.28
PCs variance % of the total variance of variables						
PC1		71.71	62.21	61.34	59.05	69.60
PC2		28.29	37.79	38.66	40.95	30.40
CV%		12.88	6.98	6.60	12.77	3.84

Statistically significant differences at * $p \leq 0.05$ and ** $p \leq 0.01$; ns: indicate the non-significant difference. LW/P: leaves weight/plant; RL/P: root length/plant; RD/P: root diameter/plant; RW/P: root weight/plant; TSS%: total soluble solids percentage %

Estimates of the environmental index under the tested environments ranged from -64.66 to 39.75, demonstrating significant differences across these environments for studied traits. The maximum values of the environmental index were recorded for LW/P (29.42), RD/P (0.17), RW/P (39.75) and TSS% (2043) traits in E2, followed by RD/P (0.05) and RW/P (24.90) in E1, indicating these environments were favorable for these studied traits. While other environments were unfavorable for the other studied traits. It is evident from the environmental index that an environment that was good for a trait was poor for another [34].

3.3 GGE Biplot Model

3.3.1 Discrimitiveness vs. Representativeness

The GEI is partitioned into two components (PC1 and PC2) by GGE biplot. The GGE biplot of PC1 and PC2 contributed 77.50% and 15.60%, and collectively they explained 93.10% of the total G+GE variation for root weight/plant (Fig. 1). After partitioning GEI by GGE biplot, the PC1 contribution in the GEI was greater than the PC2 under the various environments, suggesting that the GGE biplot effectively partitioned the variability in root weight trait. A similar trend has been reported in sugar beet by Curcic et al. [16] and Studnicki et al. [19]. Finding the most suitable (ideal) test environment through the test-environment evaluation is crucial for a successful breeding technique in the selection of superior

genotypes for a variety of environments [35]. The idealness of the tested environments is defined by two characteristics: a) discriminating ability (the ability of an environment to differentiate genotype in terms of main genotype effects), which has a high PC1 score, and b) representativeness (the ability of an environment to represent all other evaluated environments), which has a zero score for PC2. As a report by Yan and Tinker [15], the length vectors and the cosine of the angle between the two environments determine their similarity (covariance) of them. Therefore, the environments for RW/P were divided into two distinct groups by the ray's lines. The first group included the E3 environment in a sector of its own and had two suitable varieties (G3 and G9), while E1 and E2 environments formed the second group where varieties G5 and G6 had the highest RW/P. As a report by Yan et al. [35], due to having the smallest angles with average environment coordination (AEC), the test environments E1 and E2 for RW/P are more representative of other test environments (Fig. 1). These results indicate that these environments are idyllic and have the greatest ability to discriminate genotypes, thus favoring the selection of superior genotypes. Yan et al. [8] and Yan and Rajcan [36] reported that the most genotypes desirable is the one closest to the graph of the ideal environment. Thus, the genotypes G2, G5 and G6 are the most productive and stable for RW/P. While the test environment E3 had a larger angle with AEC for

RW/P, indicating that it was the least discriminating and representative in both irrigation conditions. Non-discriminating test environments provide minimal information about genotypes and must not be used as test environments [15]. According to Yan and Kang [37], strong positive correlation was observed among E1 and E2 (the acute angle), while E3 had positively correlated with E1 (slight). GGE biplot model indicated the most representative testing environments with the discriminating ability for root yield trait tested [18].

3.3.2 Mean vs. Stability analysis

The AEC method based on genotype-focused singular value partitioning (SVP) was utilized to assess genotype yield stability using average PCAs in all environments. If $SVP = 1$, the AEC line with a single arrow passes through the biplot's origin [38], the arrow points to a higher mean yield. The mean of PC1 and PC2 of the environmental scores is defined, as a report by Yan and Rajcan [36]. The 'Mean vs. stability' view is frequently referred to as AEC with $SVP = 1$ which helps to simplify the genotype assessment based on the mean performance and stability across environments within a multi-environment (Fig. 2). The GGE biplot was created by plotting the PC1 and PC2 produced from subjecting data of environment-centered yield to singular value decomposition [8]. The genotypes are grouped according to their average root weight, as indicated by the arrow sign on the AEC. The genotypes with above-average means were G2, G4, G5, and G6, while the other genotypes were below-average means. The highest values of RW/P were observed by the genotypes G5, and G6 in the E1 and E2 environments, and the genotypes G6 and G9 in the E3 environment. While G3, G7, and G10 genotypes had the least mean root weight in E2, E3, and E1, respectively. GGE biplot model revealed that some environments were the best test environments where the best genotypes could easily be identified with respect to the root trait of sugar beet [18].

The most stable genotypes were $G6 > G2 > G5 > G4$, which were practically on the AEC abscissa and had a close to zero projection onto the AEC ordinate. This demonstrates that these genotypes' ranking was remarkably constant across environments, according to Yan et al. [35]. In addition to good sugar beet root weight, genotype stability is more important, in terms of

Yan [39] established an "ideal" genotype based on average performance as well as stability. Thus, the genotypes G6 and G5 are more stable with better mean sugar beet yield other than the other genotypes. Conversely, the genotypes G1 and G3 are more variable and highly unstable with below and above-average mean performance in both conditions, respectively. The use of GGE biplots helped to indicate the mega-environments and sugar beet genotypes that yield the best in each of them [19]. Some genotypes of sugar beet had specific adaptability to the best environment identified for root yield in the GGE biplot model [18].

3.3.3 Which-won-where pattern

The polygon view of the GGE biplot pattern of root weight of sugar beet yield was constructed to show which genotypes with the best performance best in which environment and groups of environments [8], as well as to demonstrate the presence of crossover GEI, mega-environment differentiation, and specific adaptation [15]. Four genotypes including G2, G4, G8 and G9 have situated within the polygon. While other genotypes are located away from the biplot origin in all directions and which formed the polygon vertices (Fig. 3). A line perpendicular to each polygon side was drawn starting from the biplot origin. The biplot is divided into sectors by these lines. The rays are perpendicular lines to the sides of the polygon or their expansion [38]. Thus, the three environments are divided into different apparent groups. The genotype at the vertices of each sector is the nominal highest yielder for the environments or mega-environments that fell into it. Accordingly, G5 produced maximum root weight in the E1 environment. Also, the genotypes G5, G6, and G2 perform best in the E1 and E2 environments and gave the highest root weight, due to being located in the sector of these environments. While the genotype G4 occurred in the sector of the E3 environment and gave the highest root weight after genotypes G5 and G6. The genotypes G1, G3, G7, and G10 were the poorest across the environments, due to no environment falling into the sectors of these genotypes. GGE biplot analysis suggested that optimal locations were identified for each genotype, which can be useful when recommending sugar beet varieties for certain growing areas of this field crop [16]. Winner genotypes were also identified based on the results of the GGE biplot model [18].

Table 3. Mean values of studied traits of ten sugar beet genotypes grown under different environments

Traits	Environments	Genotypes										Mean	LSD at		CV%	EI
		G1	G2	G3	G4	G5	G6	G7	G8	G9	G10		0.05	0.01		
LW/P	E1	65.57	69.27	72.97	58.67	99.37	107.13	104.17	79.33	95.00	69.00	82.05	6.79	9.04	5.84	-12.76
	E2	107.43	140.00	127.60	117.93	133.33	123.70	102.87	122.33	163.50	103.50	124.22	26.42	NS	15.02	29.42
	E3	65.20	81.53	88.47	78.60	77.73	84.00	63.23	65.67	91.07	85.93	78.14	12.35	NS	11.17	-16.66
	Mean	79.40	96.93	96.34	85.07	103.48	104.94	90.09	89.11	116.52	86.14	94.80	NS	NS	12.88	
RL/P	E1	15.90	17.47	12.20	14.70	14.43	16.83	15.53	16.90	12.03	12.33	14.83	1.36	1.82	6.49	0.08
	E2	16.70	15.07	11.03	14.67	15.37	15.27	14.97	15.60	14.50	13.67	14.68	1.80	2.40	8.66	-0.07
	E3	15.63	16.33	14.03	15.07	14.60	15.17	15.00	15.30	14.07	12.27	14.75	1.12	1.50	5.38	-0.01
	Mean	16.08	16.29	12.42	14.81	14.80	15.76	15.17	15.93	13.53	12.76	14.75	0.81	1.08	6.98	
RD/P	E1	8.33	8.33	7.60	6.83	12.23	8.67	7.37	6.93	10.23	7.03	8.36	0.95	1.26	8.03	0.05
	E2	9.27	8.67	7.63	9.00	9.13	8.60	7.93	8.03	9.07	7.40	8.47	0.80	1.07	6.70	0.17
	E3	7.43	7.83	9.87	7.33	8.63	8.17	7.10	7.13	9.97	7.30	8.08	0.49	0.66	4.30	-0.23
	Mean	8.34	8.28	8.37	7.72	10.00	8.48	7.47	7.37	9.76	7.24	8.30	0.43	0.57	6.60	
RW/P	E1	460.17	762.03	582.10	781.00	1078.67	1001.27	691.67	591.67	571.00	453.33	697.29	105.23	140.14	10.66	24.90
	E2	781.23	708.33	513.33	670.00	845.33	856.30	690.20	706.67	738.33	611.67	712.14	166.96	NS	16.56	39.75
	E3	427.67	595.33	684.20	670.33	735.47	757.17	423.70	510.00	752.77	520.63	607.73	73.46	97.82	8.54	-64.66
	Mean	556.36	688.57	593.21	707.11	886.49	871.58	601.86	602.78	687.37	528.54	672.39	67.75	90.22	12.77	
TSS%	E1	16.33	19.33	16.67	16.00	15.33	17.67	16.00	17.67	16.00	16.67	16.77	0.96	1.27	4.03	-1.83
	E2	21.00	22.33	19.00	23.33	21.33	21.00	20.67	22.00	20.33	19.33	21.03	1.19	1.59	4.00	2.43
	E3	16.67	18.67	17.33	17.33	18.33	18.67	16.67	19.33	18.67	18.33	18.00	0.85	1.13	3.33	-0.60
	Mean	18.00	20.11	17.67	18.89	18.33	19.11	17.78	19.67	18.33	18.11	18.60	NS	NS	3.84	

E1, E2 and E3: Saint Catherine, Baloza and East El-Qantra, respectively. EI: Environmental index. The traits key names can be found in Table 2

3.3.4 Cluster analysis and Principle Component Analysis (PCA)

Based on the mean performances of all studied traits under this study, the hierarchical cluster analysis method was performed to classify the ten sugar beet genotypes into four clusters (Fig. 4). Each cluster contained genotypes that were highly similar. The first and fourth clusters comprised two genotypes including G5 and G6,

as well as G1 and G10, respectively. While both second and third clusters enclosed three genotypes including G2, G9 and G4, as well as G3, G7 and G8, respectively. The genotypes in the first cluster had recorded the highest root weight/plant, followed by the genotypes in the second and the third clusters, opposite is true for the genotypes in the fourth cluster. According to Hu et al. [22], cluster analysis classifies sugar beet genotypes into four clusters.

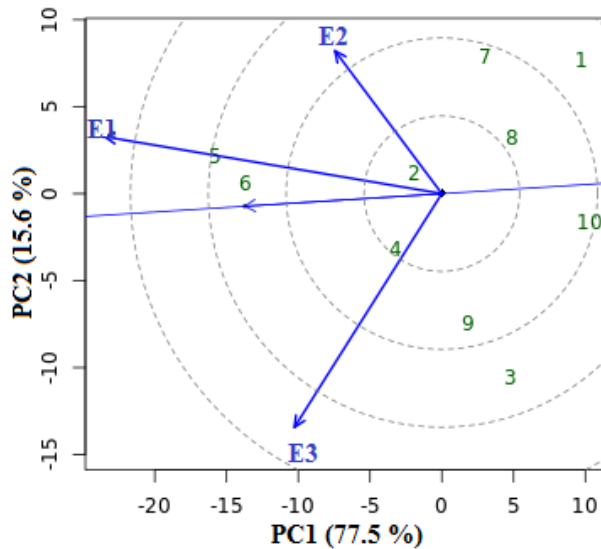


Fig. 1. GGE biplot of discriminativeness vs. representativeness for root weight/plant traits with ten sugar beet genotypes (green color) and three environments (blue color)
E1, E2 and E3: Saint Catherine, Baloza and East El-Qantra, respectively; 1: SK58-4-S4; 2: SK48-C; 3: SK70; 4: SK90; 5: SK44-1-4R2; 6: SK27-38; 7: SK27-32; 8: SK Fc723; 9: SK44-1-9C1; 10: SK17-2-7W

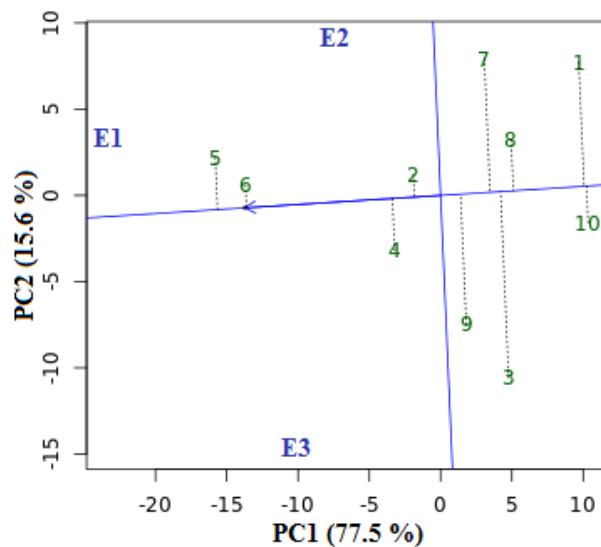


Fig. 2. GGE biplot of mean vs. stability for and root weight/plant traits with ten sugar beet genotypes (green color) and three environments (blue color). The genotypes and environment key names can be found in Fig. 1

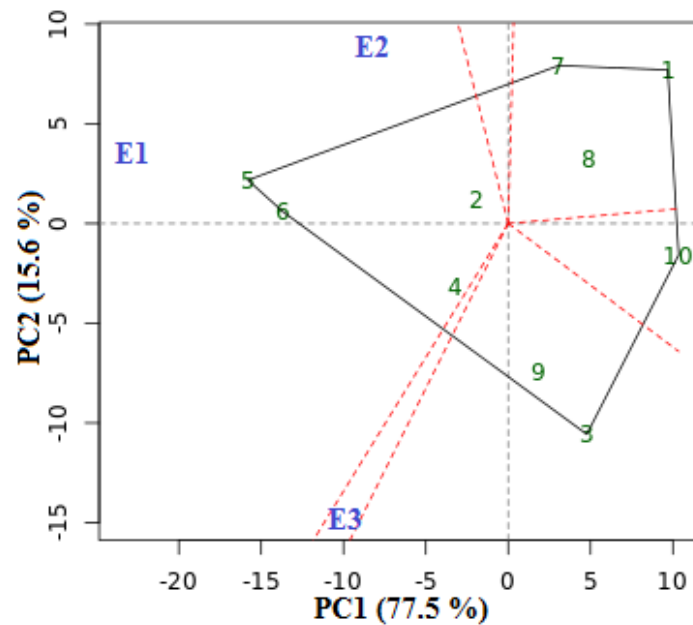


Fig. 3. GGE biplot polygon of "which-won-where" for root weight/plant with ten sugar genotypes (green color) and three environments (blue color). The genotypes and environment key names can be found in Fig. 1

To find the optimal genotypes in the investigated environments and to clearly comprehend the correlations between the studied traits, the PCA analysis was carried out for the investigated traits and genotypes. Table 4 lists the five PCs for the analyzed genotypes and traits influenced by the three environments. The first two main PCs (PC1 and PC2) extracted had eigenvalues higher than one with values of 2.35 and 1.75, respectively, and they explain 82.01% of the total variance of variables. In contrast, the other PCs have eigenvalues below one (Eigenvalue <1). The PC1 and PC2 explained 47.03%, and 34.98% of the total variance of variables, respectively. As a result, they can be used as the basis for evaluating the genotypes and the association between investigated traits under the studied environments. The PC1 had a positive correlation with all studied traits and with the genotypes G2, G5, G6 and G9. While, the PC2 is positively correlated with RL/P, RW/P and TSS% traits and with the genotypes G1, G2, G4, G6 and G8. These results indicated that PC1 can be referred to as the high-root weight component and the basis in the weighting of selection genotypes, thus PC1 is important to increase sugar beet productivity in the three environments. Alami et al. [23], Islam et al. [24], Majumdar et al. [26] and Mehareb et al. [40] have previously revealed similar results for the first two main PCs.

Based on their data from the three researched environments, the PC1 and PC2 were used to draw a biplot for the studied attributes and genotypes (Fig. 5). Using PCA, a sharp angle among LW/P, RD/P and RW/P traits, as well as among RL/P, RW/P and TSS% traits in this study was found, indicating the positive correlation between these variables, but they differed in their degree and consistency in quantity. Generally, RW/P positively correlated with the other studied traits. This means that selection based on these traits will result in an increasing sugar beet yield in all environments [31]. Evaluation of the genotypes according to their locations on the basis of the studied variables by using the PCA model might be a feasible approach [22]. The PC1 and PC2 mainly distributed and distinguished the studied traits and genotypes into two groups. The first group was related to PC1 and includes all studied traits, which are strongly positively associated with the G2, G5, G6 and G9 under the three environments (the first and fourth quarters). While the second group is related to PC2, which includes the other genotypes (the second and third quarters). The PCA with the AMMI model enables clustering of genotypes based on similarity of response characteristics and identifying potential trends in environments [41]. Generally, the G5 and G6 genotypes were located near the most studied traits across the

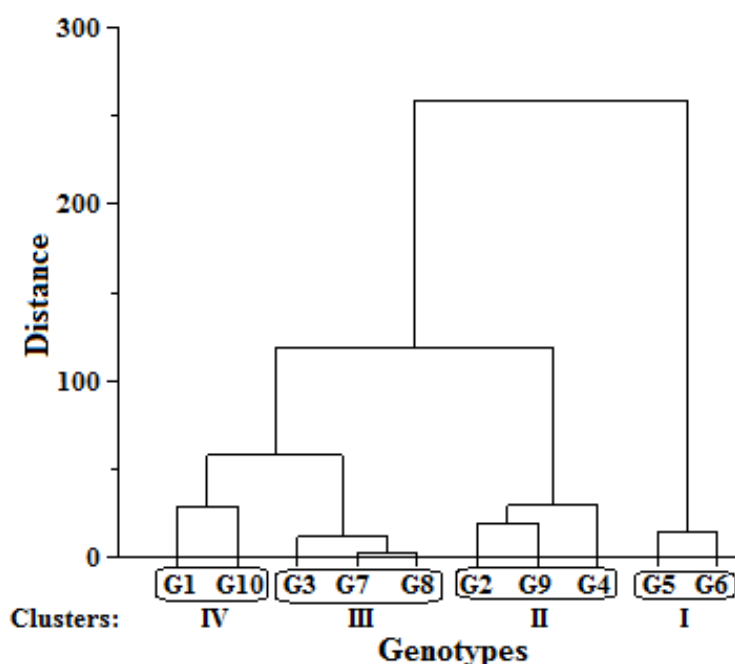


Fig. 4. Cluster dendrogram of ten sugar beet genotypes based on all studied traits across various environments. The genotypes key names can be found in Fig. 1

Table 4. Results of Principal Component analysis (PCs) in the first five PCs for the studied traits during the main effects of experimental factors

Variables	PC1	PC2	PC3	PC4	PC5
SW/P	0.58	-0.18	0.45	0.22	0.62
RL/P	0.05	0.68	-0.51	0.35	0.40
RD/P	0.57	-0.21	-0.31	0.49	-0.54
RW/P	0.57	0.18	-0.24	-0.77	-0.02
TSS%	0.14	0.65	0.62	0.07	-0.41
G1	-1.35	0.24	-1.34	0.71	-0.18
G2	0.48	1.94	0.64	0.44	-0.20
G3	-0.49	-2.03	0.34	-0.10	-0.13
G4	-0.64	0.60	-0.07	-0.67	-0.35
G5	2.41	-0.41	-0.85	-0.29	-0.38
G6	1.67	0.98	-0.02	-0.65	0.45
G7	-1.19	-0.29	-0.55	-0.06	0.76
G8	-0.95	1.63	0.60	0.22	0.02
G9	1.98	-1.47	0.61	0.76	0.16
G10	-1.91	-1.19	0.64	-0.36	-0.17
Eigenvalues	2.35	1.75	0.50	0.27	0.13
Variance %	47.03	34.98	9.97	5.40	2.62
Cumulative%	47.03	82.01	91.98	97.38	100.00

The traits and genotypes key names can be found in Table 2 and Fig. 1, respectively

three environments. The results of PCA analysis were in accord with the results of cluster analysis with respect to genotypes delineation with higher root weight and most studied traits. Based on our

results, the G5 and g6 genotypes across the three environments have the potential to improve plant growth and increase the sustainable productivity of sugar beet in Egypt.

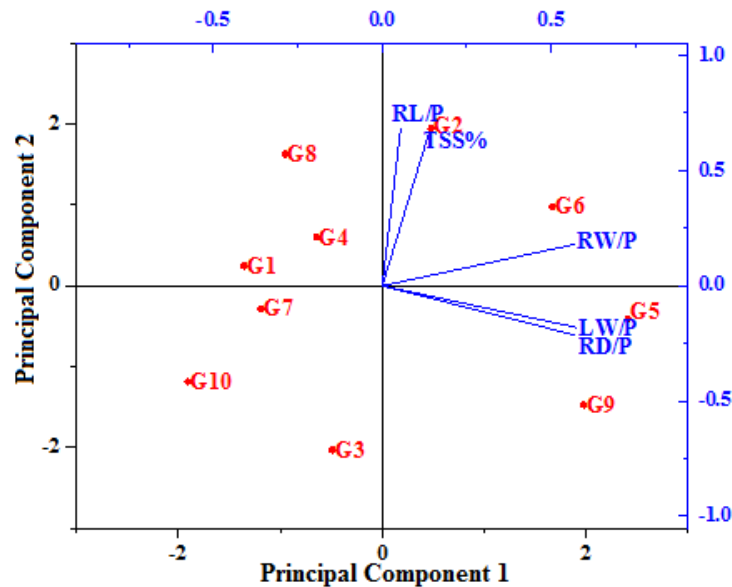


Fig. 5. A biplot diagram based on the first two PCs shows the relationships among the measured traits across ten sugar beet genotypes under various environments. The traits (blue color) and genotypes (red color) key names can be found in Table 2 and Fig. 1, respectively

4. CONCLUSIONS

The results of AMMI analysis reflect the divergent climatic conditions of three environments, resulting in a high level of genetic variability among ten genotypes for sugar beet crop under these environments. GGE biplot performed well in the study of the GEI, and provide a clear idea of genotype stability behavior in the different environments. PCA and cluster analysis could be used as suitable methods to identify the best genotypes in the examined environments and to clearly understand the association between the studied traits. The multivariate models were useful in identifying the G5 and G6 as the most stable genotypes with the greatest potential for high-yielding across various environments. Thus, these genotypes can be used in future breeding programs to higher root yields of sugar beet across different environments in Egypt.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. El-Refaey Ra, El-Seidy Eh, El-Geddawy Ih, El-Sayed Hm. Phenotypic and genotypic stability for some sugar beet genotypes.

2. Proc. 13th International Conf. Agron, Fac.of Agic, Benha Univ. Egypt. 2012;317-331.
3. Al Jbawi Ea, Al Raei F, Al Ali A, Al Zubi H. Genotype – environment interaction study in sugar beet (*Beta vulgaris* L.). International Journal of Environment. 2016; (5)3:74-86. DOI: 10.3126/ije.v5i3.15706
4. Sorour Ma, Mehanni Ae, Mahmoud Ea, Gaber Noha F. Sugar beet quality and juice purity of some sugar beet varieties (*Beta vulgaris* L.) Grown in toshka region, egypt as effected by harvesting ages and storage conditions. Archives of Agriculture Sciences Journal. 2020;3(3):64-81. DOI: 10.21608/aasj.2020.43246.1037
5. Anar Mj Lin Z, Hoogenboom G, Shelia V, Batchelor Wd, Teboh Jm, Ostlie M, Schatz Bg, Khan M. Modeling growth, development and yield of sugar beet using dssat. Agric. Syst. 2019;169:58-70. Available:https://doi.org/10.1016/j.agry.2018.11.010
6. Faostat. Food and agriculture organization of the United Nations; 2022. (Access on 18 Aug 2022). Available:https://www.fao.org/faostat/en/#d ata/qcl
7. El-hashash Ef, Hassan Ae, Agwa Am. Genotype by environment interaction effects and evaluation of drought tolerance indices under normal and severe stress

- conditions in barley. *Int J PLant Breed Crop Sci.* 2018;5:300-316.
7. Falconer Ds, Mackay Tfc. *Introduction to quantitative genetics*, 4th edition longman, New York. 1996;132-133.
 8. Yan W, Hunt La, Sheng Q, Szlavnicz Z. Cultivar evaluation and mega-environment investigation based on gge biplot. *Crop sci.* 2000;40:596-605.
Available:<https://doi.org/10.2135/cropsci2000.403597x>
 9. Björnsson J. *Stability analysis towards understanding genotype x environment interaction..* Plant agriculture department of university of guelph, ontario, Canada; 2002.
(Access on 18 Aug 2022)
Available:www.Genfys.Slue.se/staff/deg/nova02
 10. Ssemakula G, Dixon A. Genotype x environment interaction, stability and agronomic performance of carotenoid-rich cassava clones. *Sci res essays* 2007;2: 390–399.
Available:<https://cgspace.cgiar.org/handle/10568/92828>
 11. Hoffmann Cm, Huijbregts T, Van Swaaij N, Jansen R. Impact of different environments in europe on yield and quality of sugar beet genotypes. *European Journal of Agronomy.* 2009;30:17-26.
Available:<https://dx.doi.org/10.1016/j.eja.2008.06.004>
 12. Gauch Hg, Piepho Hp, Annicchiarico P. Statistical analysis of yield trials by ammi and gge: further considerations. *Crop Sci.* 2008;48:866–889.
DOI: 10.2135/cropsci2007.09.0513
 13. Zobel Rw, Wright Mg, Gauch Hg. Statistical analysis of yield trial. *Agron. J.* 1988;80(3):388-393.
Available:<https://doi.org/10.2134/agronj1988.00021962008000030002x>
 14. Gabriel Kr. The biplot graphic display of matrices with applications to principal component analysis. *Biometrics.* 1971;58: 453-467.
Available:<https://doi.org/10.2307/2334381>
 15. Yan W, Tinker Na. Biplot analysis of multi-environment trial data: principles and applications. *Can J Plant Sci.* 2006;86: 623-645.
Available:<https://doi.org/10.4141/p05-169>
 16. Curcic Z, Danojević D, Bojan M, Ciric M, Taski-Ajdukovic K, Nagl N. Gge biplot analysis of sugar beet multi-environment trials. *Povrtarstvo.* 2017; 54(2):61-67.
DOI: 10.5937/ratpov54-13241
 17. Curcic Z, Ciric M, Nagl N, Taski-Ajdukovic K. Effect of sugar beet genotype, planting and harvesting dates and their interaction on sugar yield. *Front. Plant Sci.* 2018; 9:1041.
DOI: 10.3389/fpls.2018.01041
 18. Hassani M, Heidari B, Dadkhodaie A, Stevanato P. Genotype by environment interaction components underlying variations in root, sugar and white sugar yield in sugar beet (*Beta vulgaris* L.). *Euphytica.* 2018;214:79.
Available:<https://doi.org/10.1007/s10681-018-2160-0>
 19. Studnicki M, Lenartowicz T, Noras K, Wójcik-Gront E, Wyszynski Z. Assessment of stability and adaptation patterns of white sugar yield from sugar beet cultivars in temperate climate environments. *Agronomy.* 2019;9(7):405.
Available:<https://doi.org/10.3390/agronomy9070405>
 20. Bocianowski J, Wielkopolan B, Jakubowska M. Ammi analysis of the effects of different insecticidal treatments against agrotis spp. On the technological yield from sugar beet. *Agriculture.* 2022a; 12:157.
Available:<https://doi.org/10.3390/agriculture12020157>
 21. Škrbić B, Đurišić-Mladenović N, Mačvanin N. Determination of metal contents in sugar beet (*Beta vulgaris*) and its products: Empirical and chemometrical approach. *Food science and technology research.* 2010;16(2):123-134.
Available:<https://doi.org/10.3136/fstr.16.123>
 22. Hu X-H, Zhou J-C, Yang H-Z (2019). Comprehensive evaluation of different sugar beet varieties by using principal component and cluster analyses. *Iop Conf. Series: Journal of Physics: Conf. Series.* 2019;117:1-14.
DOI: 10.1088/1742-6596/1176/4/042021
 23. Alami I, Terouzi W, Otmani M, Abdelkhalek O, Salmaoui S, Mbarki M. Effect of sugar beet harvest date on its technological quality parameters by exploratory analysis". *Journal of Food Quality.* 2021;8.

- Available:<https://doi.org/10.1155/2021/6639612>
24. Islam Mj, Uddin Mj, Hossain Ma, Henry R, Begum Mk, Sohel M, Mou Ma, Ahn J, Cheong Ej, Lim Ys. Exogenous putrescine attenuates the negative impact of drought stress by modulating physio-biochemical traits and gene expression in sugar beet (*Beta vulgaris* L.). Plos One 2022;17(1): e0262099.
Available:<https://doi.org/10.1371/journal.pone.0262099>
 25. Kleuker G, Hoffmann Cm. Causes of different tissue strength, changes during storage and effect on the storability of sugar beet genotypes. Postharvest Biology and Technology 2022;183:111744.
Available:<https://doi.org/10.1016/j.postharvbio.2021.111744>
 26. Majumdar R, Strausbaugh Ca, Vincill Ed, Eujayl I, Galewski Pj. Leaf bacteriome in sugar beet shows differential response against beet curly top virus during resistant and susceptible interactions. International Journal of Molecular Sciences. 2022; 23(15):8073.
Available:<https://doi.org/10.3390/ijms23158073>
 27. Steel Rgd, Torrie Jh. Principles and procedures of statistics. 2nd edition. Mcgraw hill book company inc., New York; 1980.
 28. Basford Ke, Cooper M. Genotype x environmental interactions and some considerations of their implications for wheat breeding in australia. Australian Journal of Agricultural Research. 1998; 49(2):153-174.
Available:<https://doi.org/10.1071/a97035>
 29. Mather K, Jinks Jl. Introduction to biometrical genetics, chapman and hall limited, london; 1971.
Available:<https://doi.org/10.1007/978-1-4613-3387-6>
 30. El-hashash Ef, Agwa Am. Comparison of parametric stability statistics for grain yield in barley under different drought stress severities. Merit Research Journal of Agricultural Science and Soil Sciences. 2018;6(7):098-111.
 31. Bayomi Kem, El-hashash Ef. Moustafa Esa. Comparison of genetic parameters in non-segregating and segregating populations of sugar beet in egypt. Asian Journal of Research in Crop Science. 2019;3(4):1-12.
Available:<https://doi.org/10.9734/ajrcs/2019/v3i430055>
 32. Yan W, Hunt La. Interpretation of genotype x environment interaction for winter wheat yield in ontario. Crop sci. 2001;41(1):19-25.
Available:<https://doi.org/10.2135/cropsci2001.41119x>
 33. Thillainathan M, Fernandez Gcj. A novel approach to plant genotypic classification in multi-site evaluation. Hort. Sci. 2002; 37(5):793-798.
Available:<https://doi.org/10.21273/hortsci.37.5.793>
 34. El-Hashash Ef, Km El-absy Km. Genotype x environment interaction, environmental indices and stability analyses for some selected genotypes of barley. Journal of Applied Sciences Research. 2013;9(7): 4371-4377.
 35. Yan W, Kang Ms, Ma, Bl, Woods S, Cornelius Pl. Gge biplot vs. Ammi analysis of genotype-by-environment data. Crop sci. 2007;47(2):643-655.
Available:<https://doi.org/10.2135/cropsci2006.06.0374>
 36. Yan w, rajcan I. Biplot analysis of test sites and trait relations of soybean in ontario. Crop sci. 2002;2(1):11-20.
Available:<https://doi.org/10.2135/cropsci2002.1100>
 37. Yan W, Kang Ms. GGE biplot analysis: A graphical tool for breeders, geneticists, and agronomists (1st Ed.). Crc press. 2002.
Available:<https://doi.org/10.1201/9781420040371>
 38. Yan W. Singular-value partition for biplot analysis of multi-environment trial data. Agronomy J., 2002;94(5):990-996.
Available:<https://doi.org/10.2134/agronj2002.9900>
 39. Yan W. Ggebiplot-a windows application for graphical analysis of multi-environment trial data and other types of two-way data. Agron. J. 2001;93(5):1111-1118.
Available:<https://doi.org/10.2134/agronj2001.9351111x>
 40. Mehareb Em, El-bakary Hmy, Abo Elenen Ffm. Comprehensive evaluation of sugar beet genotypes for yield and relative traits by multivariate analysis. Svui-international Journal of Agricultural Sciences. 2021;3(1): 96-111.
DOI: 10.21608/svuijas.2021.58999.1072

41. Bocianowski J, Jakubowska M, Kowalska J. The interaction of different abiotic conditions on the value of the component traits of the technological yield of sugar beet. *Euphytica*. 2022b;218: 110. Available:<https://doi.org/10.1007/s10681-022-03070-4>.

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