

Research Article

Untargeted Metabolite Profiling of Specialty Rice Grains Using Gas Chromatography Mass Spectrometry

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With-ever increasing demand food grains for the increasing population, it has also increased the importance of quality rice with nutritional and therapeutic properties. The quality of rice includes nutritional value, therapeutic properties, and further generation of aroma. Initial studies on sensory analysis using potassium hydroxide (1.7% KOH) identified the presence of a distinct aroma of the traditional rice cultivar Chakhao Amubi in comparison with other aromatic rice varieties were conducted. The metabolomic profiling of aromatic rice grains Chakhao Amubi, Pusa Basmati 1, and nonaromatic rice, Improved White Ponni was attempted to use gas chromatography-mass spectrometry (GC-MS). A total of fifty volatile aromatic compounds, including aromatic hydrocarbons, alkanes, alkenes, ketones, and aromatic aldehydes, have been identified. Detected compounds include six crucial volatile i.e., pentanal, hexanal, 2-pentylfuran, pyridine, (Z)-7-Decenal, and Mesitylene for distinct flavor and presence of aroma in Chakhao Amubi. The findings showed a distinct difference in the metabolic profile of Chakhao Amubi compared to Pusa Basmati 1 and Improved White Ponni. Thus, this study paved the way for a new understanding of the aromatic aspects of traditional rice germplasm and its utilization in rice breeding programs to improve the aroma, therapeutic, and nutritional characteristics of rice.

1. Introduction

Being a staple food for more than half of the world's population, rice holds a key position in achieving food security. It is predicted that the global population will reach beyond 9.7 billion in 2050, which necessitates a doubling of rice production to ensure food and nutritional security [1]. Even though rice serves as a staple food for the majority of the population, its consumer preference varies greatly between different countries. In India, preference for rice quality (size, taste, and aroma) differs between regions [2]. Next to the grain size and cooking quality, aromatic rice is highly preferred by the majority of consumers because of the fragrance and its unique taste. Aromatic rice is considered to be the most precious asset of rice diversity to India and fetches a premium price (3 times higher price than non-aromatic rice) in the export market. Among the various fragrant rice cultivars, Sadri (Iran), Basmati (India and Pakistan), Jasmine (*indica* rice from Thailand) and a few

tropical *japonicas* are ruling the market [3]. Systematic screening of >250 volatile and nonvolatile compounds in aromatic and nonaromatic rice varieties identified a compound, 2-acetyl-pyrroline (2AP), majorly responsible for the production of pop-corn like aroma in rice [4, 5]. Aromatic rice varieties greatly differ in their 2AP content, as reported in Basmati (0.34 ppm), Jasmine (0.81 ppm), and Texmati (0.53 ppm). Later, it was found that 2AP has present in both aromatic and nonaromatic rice varieties, but they differed in concentration [6]. In addition to 2-acetyl-1-pyrroline, other novel compounds like 2-amino acetophenone and 3-hydroxy- 4, 5-dimethyl-2 (5H) -furanone in Basmati 370 [7] and guaiacol and indole and p-xylene in Black rice [8] were also reported to contribute for a unique flavor in the rice. In addition to 2-acetyl-1-pyrroline, more than 100 volatile organic compounds, namely, hexanal, nonanal, octanal, nonenal, (E, E)-2, 4-nonadienal, heptanal, (E)-2-octenal, 4-vinyl phenol, 4-vinyl guaiacol, 1-octen-3-ol, decanal, have been reported. [9, 10].

Recent advancements in molecular genetics and sequencing enabled the molecular dissection of aroma in rice. QTL mapping and comparative sequence analysis of OsBADH2 gene on chromosome 8 between the aromatic rice KDML105 and a nonaromatic rice Nipponbare identified key mutations in the exon 7 of OsBADH2 namely, two base pair substitutions, one at 730 bp (A to T), another base pair substitution at 732 bp (T to A), and an 8-bp deletion "GATTAGGC" starting from 734 bp [11]. Similar studies in the aromatic rice variety Kayeema also identified an 8 bp deletion in the exon 7 of OsBADH2 [12, 13]. In addition to the fragrance gene OsBADH2 on chromosome 8, several other minor QTLs located on chromosomes 3, 4, and 12 have a strong associations with aroma reported [11]. In a similar manner, comparative aroma profiling in rice identified novel aromatic compounds like alkanals, alk-2-enals, alka (E)-2, 4-dienals, 2-pentylfuran, 2-acetyl-1-pyrroline, and 2-phenylethanol.

India has a huge asset of different aromatic rice in both Basmati (Group V) and *indica* gene pools. There are regional specific aromatic rice, namely, Basmati (UP, Punjab, Bihar, and Haryana), Kalanamak (Eastern UP), Seeraga samba (Tamil Nadu), Jeerasala/Ganda sala (Kerala), and Chakhao Amubi/Chakhao Poireiton (North East India). Genomic analysis of Seeraga samba identified a novel mutation in OsBADH2 leading to aroma [14]. Sensory evaluation of a few of the above aromatic rice grains revealed the presence of a distinct aroma when compared to 2AP possessing Basmati (unpublished data). When compared to measuring the allelic diversity of OsBADH2, only a few attempts have been made in profiling aromatic compounds with fragrance traits in rice. Identification of novel aromatic compounds contributing to fragrance in rice and subsequent genetic analysis will accelerate the development of high-yielding fragrant rice varieties for the export market. In this study, attempts were made to profile the aromatic volatile compounds in three contrasting rice genotypes, namely, Improved White Ponni (nonaromatic fine grain variety), Pusa Basmati (aromatic long slender rice), and Chakhao Amubi (aromatic black rice from North East India).

2. Materials and Methods

2.1. Genetic Materials Used. Three different rice genotypes, namely, Improved White Ponni (nonaromatic indica), Chakhao Amubi (aromatic black rice of North East India), and Pusa Basmati 1 (aromatic long slender basmati) were used in this study. Sensory evaluation test was used as reported by Hien et al. (2006) 1.7 percent potassium hydroxide (KOH) for analyzing scent from grains of Chakhao Amubi and Pusa Basmati 1. One gram of dehusked seeds was powdered using a mortar and pestle. The samples were transferred to petri dishes containing 10 mL of 1.7% potassium hydroxide solution. The petri dishes were closed and retained at room temperature for 10 min. After 10 minutes, the dishes were opened and smelled. The presence or absence of aroma was scored (Hien et al., 2006). A sensory evaluation tests in the grains of the above 3 rice genotypes revealed that the aroma in the grains of Chakhao Amubi differed strikingly from Pusa Basmati 1 (unpublished data).

2.2. Profiling Volatiles in the Grains of Contrasting Rice Genotypes through GC-MS

2.2.1. Sample Preparation and Volatile Capturing. Freshly harvested, dehusked grains of Chakhao Amubi, Pusa Basmati 1, and Improved White Ponni was used. 10 g dehusked seeds were ground into powder using a clean blender. Fine rice flour was transferred to a sterile conical flask containing 100 ml of 1.7% potassium hydroxide. The conical flask was tightly closed with a rubber cap containing a collector tube and sealed with a parafilm wrap to avoid any leakage. For extraction and collection of volatiles, the powered rice of three genotypes was exposed separately using 1.7% potassium hydroxide, and released volatiles were captured using stainless steel ATD prepackaged sample collection tubes containing Sorbent: Tenax GR. The conical flask containing the sample was kept at room temperature (27–30°C) overnight (8 hours) to facilitate the collection of the volatiles in the collector tube. Sorbent tubes containing trapped volatiles were removed from the flask and directly fed into the GC-MS.

2.3. GC-MS Analysis. Gas Chromatography-Mass Spectrometry (GC-MS) was performed using a Clarus SQ 8C GC/Mass Spectrometer (Perkin Elmer, USA). Nonpolar, standard, DB-5 ms capillary columns with a dimension of 30 Mts X 0.25 um were used. Helium (He) gas was used as a carrier. The temperature program was set at 75° C and ramped to 260° C at 10°C/min, with a hold for 2 min. The source and transfer temperatures were set to 220°C and 250°C, respectively. Identification of compounds was performed by data acquisition and evaluation using the PerkinElmer Turbo Mass software Ver6.1.0. The GC-MS analysis was obtained by matching the retention time of mass spectra and compared with the reference mass spectra in the NIST library database.

3. Results and Discussion

Rice is a major staple food for more than 50% of the human population and its preference is determined by grain size, cooking quality, and aroma. Next to its grain size and cooking quality, aroma or fragrance in rice grains is a key trait determining consumer preference and market price. Several experiments have concluded that aroma in rice is determined by 1-(3, 4-dihydro-2H-pyrrol-5-yl) ethanone commonly known as 2-acetyl-1-pyrroline (2AP) [9, 15]. Production and accumulation of 2AP in rice are attributed to loss-of-function mutations in the coding region of the fragrance gene, namely, OsBADH2 (Os08g0424500) [12, 16]. Any interference in the expression of OsBADH2 in nonaromatic rice led to an accumulation of 2AP and thereby enhanced aroma [17, 18] and it is overexpression reduced the level of aroma [19]. Independent genetic studies have identified multiple alleles that have been found in the same FGR gene in different genetic backgrounds and the level of aroma differs between varieties.

India is a home for scented rice varieties, and ancient records documented the cultivation of more than 300

aromatic rice varieties in various states of India [20]. Some of the regional specific aromatic rice's include Basmati (North West India), Kalanamak (Uttar Pradesh), Dubraj and Chinnor (Madhya Pradesh), Ambemohor (Maharashtra), Radhunipagla (West Bengal), Jeeragasamba (Tamil Nadu), Gandhakasala (Kerala), Kalajira (Odisha), and Chakhao Amubi and Chakhao angouba (Manipur). Scented rice varieties of India differ greatly in their grain size and aroma. The majority of the scented rice is thermo/photo sensitive and short-grained and a few of them or long-grained. Aromatic and nonaromatic rice were classified mainly based on the content of 2 AP. Studies at IIRR, Hyderabad identified a few aromatic rice genotypes like Tarenbogh, Bansphool, Adamchini, and Ganjeikalli with no deletion in OsBADH2 and possessing no 2AP [21]. Ancient literature described five distinct types of scented rice, including mahasali, sugandhika, promodhaka, and pundrika. Grouping of aromatic rice genotypes based on isozyme banding pattern detected six different groups of scented rice [22], Similarly, previous studies have reported varying types of gene actions including monogenic, digenic, and trigenic interactions controlling aroma in rice. This indicated the existence of allelic, genic, and metabolomic diversity underlying fragrance in rice.

Profiling volatiles and semi-volatiles in aromatic rice grains detected more than 500 compounds [23]. GC-MS and LC-MS tools are becoming powerful means of profiling volatiles and metabolites. Recent advancements in sampling, collection of volatiles, and mass spectrometry identification have detected several flavor components in rice [24]. More than 40 compounds based on their mass spectra and temperature-programmed retention indices have been identified. Since 1983, 2-acetyl-1-pyrroline has been regarded as the solely most important compound responsible for the aroma of rice [4]. But, a few of the aromatic rice varieties, namely, Kao Dok MAli 105/Thai Jasmine rice and Mentik Wangi, were found to possess other aromatic compounds, namely, hexanal and 2-pentylfuran.

Phylogenetic analysis using isozyme profiles revealed that aromatic rice belongs to a distinct (Group V) cluster [22, 25]. The most popular aromatic rice includes Basmati rice from India/Pakistan, jasmine rice from Thailand, and Sadri rice from Iran. Apart from this, as discussed earlier, a few of the regional specific cultivars of India have a distinct aroma and metabolic signatures. North-Eastern India harbors greater diversity of >10,000 diverse rice cultivars including both aromatic and nonaromatic rice [26, 27]. Among them, the Chakhao cultivars of Manipur are known for their pleasant aroma and several medicinal values [28]. Few studies have attempted to analyze the metabolite diversity of different Chakhao rice types in North East India [29, 30] but no studies have attempted to profile their aroma.

Our preliminary studies through sensory evaluation tests revealed the presence of a distinctive aroma/flavor in Chakhao Amubi in comparison with other aromatic rice varieties, including Basmati. This study was aimed at profiling volatile fractions of Chakhao Amubi along with Basmati (Pusa Basmati 1) and nonaromatic rice Improved White Ponni using thermal desorption gas chromatography-mass spectrometry.

3.1. Volatiles of Rice Grains Differing in Their Aroma. GC-MS analysis of aromatic and nonaromatic rice genotypes identified more than 50 volatile compounds (Figure 1; Table 1). Out of these 50 compounds, about 10 compounds including Hexanal, o-Xylene, 2-pentylfuran, Nonanal, Undecane, 4, 7-dimethyl-cedrene, and Dodecane, were found in all three rice genotypes (Table 1). 2-pentylfuran is found to have a nutty bean odour and is also found in various food substances like alcoholic beverages, coffee, potatoes, tomatoes, and soybean oil. It imparts a floral, fruity, nutty, and caramel-like aroma to rice [31]. 2-pentylfuran was reported as the key volatile compound in jasmine rice and Mentik Wangi [42]. In many food additives, 2-pentylfuran is used as an aromatic agent. Benzaldehyde also provides a sweet, nutty flavor [31]. Hexanal is reported to be formed from the degradation of lipid products [31] and reported to emit a green grassy odour and is associated with off-flavor in rice [4]. Dimethyl trisulfide emits a sulfury, cabbage-like odour.

3.2. Pusa Basmati and Chakhao Amubi Differ in Their Aroma Profile. There were around 30 volatile compounds specific to the aromatic genotypes Pusa Basmati 1 and Chakhao Amubi (Table 1). Among them, 11 volatiles including 2AP were found to be specifically present in Pusa Basmati 1 (Table 1). Volatiles, namely, Nonane, 1-Pentanol, octanoic acid, 2-methyl-Dimethyl trisulfide, Benzaldehyde, 2-hydroxy-acetic acid, octyl ester Dodecane, Isolongifolene-5-ol, pentadecane, pentanal, and 2 Acetyl-1 pyrroline were found to be specifically present in the grains of Pusa Basmati. These results clearly indicated the presence of 2AP (C₆H₉NO), a potent aromatic compound along with other volatiles present only in aromatic Pusa Basmati 1 and not observed in Chakhao Amubi and Improved White Ponni. Several researchers have previously reported 2-acetyl-1-pyrroline as the main aromatic compound in rice [15, 31, 44], *P. Amaryllifolius* [47], sorghum [44], vegetable soybean [48], and cereal coffee brew [49].

Grains of a nonbasmati rice Chakhao Amubi native of North East India were found to possess several volatiles, namely, Toluene, p-Xylene, 1,2-Dimethylbenzene, Mesitylene, Decane, Octanal, Undecane, 2-pentynal, 2-hydroxy-Benzaldehyde, Ethyl Acetate, and beta-elemene. Interestingly, the promising candidate 2AP was absent in the grains of Chakhao Amubi. Volatile compounds detected in the grains of Chakhao Amubi have been reported earlier in other aromatic rice varieties [32–35, 50].

3.3. Chakhao Amubi Possess Novel Aromatic and Therapeutic Compounds. Generally, black rice is believed to be rich in nutraceutical compounds and hence, they were considered medicinal rice. Manipuri Chakhao rice genotypes have been assigned geographical indication (GI) status by the Government of India (GoI, 2019). Very few studies have been carried out to document the antioxidant, anthocyanin, phenolic compounds, and few other secondary metabolites of Chakhao rice genotypes [29, 30, 51]. In this study, GC-MS analysis of the grains of Chakhao Amubi identified novel

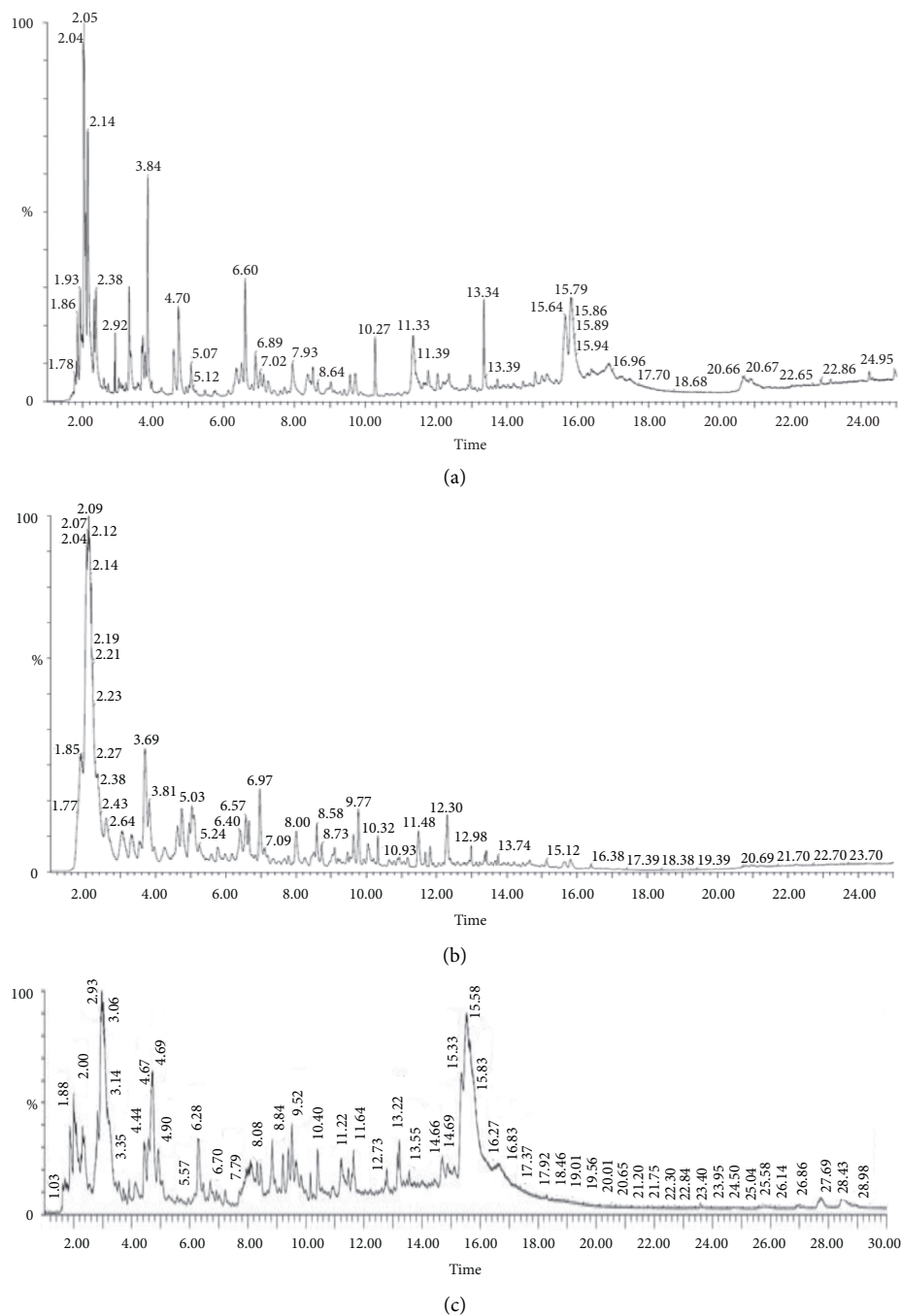


FIGURE 1: Chromatographs of (a) Chakhao Amubi (b) Improved White Ponni, and (c) Pusa Basmati 1.

therapeutic compounds, namely, Mesitylene and β -elemene which are not present in the other aromatic (Pusa Basmati 1) and nonaromatic (Improved White Ponni). Mesitylene is a colorless liquid with a sweet aromatic odor. It is found to be present in several plants, including white tea [19].

Elemenes are natural chemical compounds found in a wide variety of plants [52]. The elemenes, consisting of α -, β -, γ -, and δ -elemene, are structural isomers and contribute to the aromas of some plants and are used as pheromones by some insects. β -elemene is a primary component of

Curcuma wenyujin, exhibiting antitumor activity [53]. β -elemene is transformed from its precursor germacrene A, which is synthesized by germacrene A synthase (GAS). Our study has identified the biosynthesis of β -elemene from a cereal crop for the first time. Further studies are required to unravel the bio-synthetic pathway leading to the accumulation of β -elemene Chakhao Amubi, which will in turn pave way for introducing this novel trait into other popular rice varieties through marker-assisted breeding and metabolic engineering.

TABLE 1: List of volatile compounds identified in grains of Chakhao Amubi, Improved White Ponni, and Pusa Basmati 1 cultivars.

Group	Volatile aromatic compound	Reference
Compounds detected in the grains of all the three genotypes (Chakhao Amubi, Pusa Basmati 1, and Improved White Ponni)	Hexanal, o-Xylene, 2-pentylfuran, nonanal, undecane, 4,7-dimethyl-cedrene, and dodecane	[31–39]
Compounds detected in the grains of Pusa Basmati 1 alone	Nonane; 1-pentanol; octanoic acid; 2-methyl-, dimethyl trisulfide; benzaldehyde; 2-hydroxy-, acetic acid; octyl ester dodecane; isolongifolene-5-ol; pentadecane; pentanal, and 2 acetylc-1 pyrroline (2AP)	[31, 32, 34–41]
Compounds detected in the grains of Chakhao Amubi alone	Toluene; p-xylene; 1,2-Dimethylbenzene; mesitylene; decane; octanal; undecane; 2-pentynal; 2-hydroxy-benzaldehyde Ethyl acetate; and beta-elemene	[23, 31, 32, 34–39, 42, 43]
Compounds common to both Chakhao Amubi and Pusa Basmati	Benzaldehyde; (Z)-7-decenal; and tetradecane	[34, 35, 37–40, 42, 44]
Compounds identified in Chakhao Amubi and Improved White Ponni	2-Methylfuran; m-xylene; decanal; pentadecane	[31, 34, 36–40, 43, 45]
Compounds identified in Pusa Basmati and Improved White Ponni	Benzothiazole	[32, 37, 39, 46]

4. Conclusion

The present study revealed key differences in the volatile profile between different aromatic and nonaromatic rice varieties. This study has documented more than fifty volatile compounds in three different rice varieties differing in their aroma (Table 1). As reported earlier, 2AP was found to be the principal volatile responsible for the fragrance in Pusa Basmati 1. The delicacy of North East India, Chakhao Amubi was found to contain novel aromatic compounds, namely, pentanal, hexanal, 2-pentylfuran, pyridine, (Z)-7-Decenal, Mesitylene, and β -elemene which may be responsible for its distinct flavor and therapeutic clues. Overall, this study has identified a rice genotype, Chakhao Amubi, having the ability to accumulate or synthesize novel aromatic compounds, which will enable the molecular geneticist to understand the genetic and molecular basis of these useful properties. Above all, this study has detected Chankhao Amubi possessing a potential therapeutic compound β -elemene which needs further investigation. The present study identified valuable volatile aromatic compounds in the grains of Chakhao Amubi, which require further quantification. Identification of genes and pathways involved in the synthesis of these volatiles can be further investigated.

Data Availability

No data were used to support this study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] E. G. N. Mbanjo, T. Kretschmar, H. Jones et al., “The genetic basis and nutritional benefits of pigmented rice grain,” *Frontiers in Genetics*, vol. 11, p. 229, 2020.
- [2] A. Nayak, D. Chaudhury, and J. Reddy, “Inter-relationship among quality characters in scented rice,” *Indian Journal of Agricultural Research*, vol. 37, no. 2, pp. 124–127, 2003.
- [3] R. Singh, P. Gautam, S. Saxena, and S. Singh, *Scented Rice Germplasm: Conservation, Evaluation and Utilization*, pp. 107–133, Aromatic rices Kalyani, New Delhi, India, 2000.
- [4] R. G. Buttery, L. C. Ling, B. O. Juliano, and J. G. Turnbaugh, “Cooked rice aroma and 2-acetyl-1-pyrroline,” *Journal of Agricultural and Food Chemistry*, vol. 31, no. 4, pp. 823–826, 1983.
- [5] C. F. Lin, T. C. Y. Hsieh, and B. J. Hoff, “Identification and Quantification of the “Popcorn”-like aroma in Louisiana aromatic Delia rice (*Oryza sativa*, L.),” *Journal of Food Science*, vol. 55, no. 5, pp. 1466–1467, 1990.
- [6] U. Tanchotikul and T. C. Y. Hsieh, “An improved method for quantification of 2-acetyl-1-pyrroline, a “popcorn”-like aroma, in aromatic rice by high-resolution gas chromatography/mass spectrometry/selected ion monitoring,” *Journal of Agricultural and Food Chemistry*, vol. 39, no. 5, pp. 944–947, 1991.
- [7] M. Jezussek, B. O. Juliano, and P. Schieberle, “Comparison of key aroma compounds in cooked brown rice varieties based on aroma extract dilution analyses,” *Journal of Agricultural and Food Chemistry*, vol. 50, no. 5, pp. 1101–1105, 2002.
- [8] D. S. Yang, K.-S. Lee, O.-Y. Jeong, K.-J. Kim, and S. J. Kays, “Characterization of volatile aroma compounds in cooked black rice,” *Journal of Agricultural and Food Chemistry*, vol. 56, no. 1, pp. 235–240, 2008.
- [9] S. Mathure, A. Shaikh, N. Renuka et al., “Characterisation of aromatic rice (*Oryza sativa* L.) germplasm and correlation between their agronomic and quality traits,” *Euphytica*, vol. 179, no. 2, pp. 237–246, 2011.
- [10] A. Nadaf, S. Mathure, and N. Jawali, “Scented rice (*Oryza sativa* L.) cultivars of india: a perspective on quality and diversity,” 2016, <https://link.springer.com/book/10.1007/978-81-322-2665-9>.
- [11] Y. Amarawathi, R. Singh, A. K. Singh et al., “Mapping of quantitative trait loci for basmati quality traits in rice (*Oryza sativa* L.),” *Molecular Breeding*, vol. 21, no. 1, pp. 49–65, 2007.
- [12] L. M. T. Bradbury, R. J. Henry, Q. Jin, R. F. Reinke, and D. L. Waters, “A perfect marker for fragrance genotyping in rice,” *Molecular Breeding*, vol. 16, no. 4, pp. 279–283, 2005.
- [13] K. Sakthivel, N. Shobha Rani, M. K. Pandey et al., “Development of a simple functional marker for fragrance in rice and its validation in Indian Basmati and non-Basmati fragrant rice varieties,” *Molecular Breeding*, vol. 24, no. 2, pp. 185–190, 2009.

- [14] G. Bindusree, P. Natarajan, S. Kalva, and P. Madasamy, "Whole genome sequencing of *Oryza sativa* L. cv. Seeragasamba identifies a new fragrance allele in rice," *PLoS One*, vol. 12, no. 11, 2017.
- [15] R. G. Buttery, J. G. Turnbaugh, and L. C. Ling, "Contribution of volatiles to rice aroma," *Journal of Agricultural and Food Chemistry*, vol. 36, no. 5, pp. 1006–1009, 1988.
- [16] M. J. Kovach, M. N. Calingacion, M. A. Fitzgerald, and S. R. McCouch, "The origin and evolution of fragrance in rice (*Oryza sativa* L.)," *Proceedings of the National Academy of Sciences*, vol. 106, no. 34, pp. 14444–14449, 2009.
- [17] A. Vanavichit and T. Yoshihashi, "Molecular aspects of fragrance and aroma in rice," *Advances in Botanical Research*, vol. 56, pp. 49–73, 2010.
- [18] S. Ashokkumar, D. Jaganathan, V. Ramanathan et al., "Creation of novel alleles of fragrance gene OsBADH2 in rice through CRISPR/Cas9 mediated gene editing," *PLoS One*, vol. 15, no. 8, 2020.
- [19] S. Chen, Y. Yang, W. Shi et al., "Badh2, encoding betaine aldehyde dehydrogenase, inhibits the biosynthesis of 2-acetyl-1-pyrroline, a major component in rice fragrance," *The Plant Cell Online*, vol. 20, no. 7, pp. 1850–1861, 2008.
- [20] U. Ahuja, S. Ahuja, R. Thakrar, and N. S. Rani, "Scented rices of India," *Asian Agri-History*, vol. 12, no. 4, pp. 267–283, 2008.
- [21] K. Sakthivel, R. Sundaram, N. Shobha Rani, S. Balachandran, and C. Neeraja, "Genetic and molecular basis of fragrance in rice," *Biotechnology Advances*, vol. 27, no. 4, pp. 468–473, 2009.
- [22] J.-C. Glaszmann, "Isozymes and classification of Asian rice varieties," *Theoretical and Applied Genetics*, vol. 74, no. 1, pp. 21–30, 1987.
- [23] D. K. Verma and P. P. Srivastav, "A paradigm of volatile aroma compounds in rice and their product with extraction and identification methods: a comprehensive review," *Food Research International*, vol. 130, 2020.
- [24] M. Zeng, L. Zhang, Z. He et al., "Determination of flavor components of rice bran by GC-MS and chemometrics," *Analytical Methods*, vol. 4, no. 2, pp. 539–545, 2012.
- [25] G. Khush, "Taxonomy and origin of rice," 2000, <https://www.sciencedirect.com/science/article/pii/B9780128115084000010>.
- [26] B. Das, S. Sengupta, S. K. Parida et al., "Genetic diversity and population structure of rice landraces from Eastern and North Eastern States of India," *BMC Genetics*, vol. 14, no. 1, pp. 71–14, 2013.
- [27] A. Mao, T. Hynniewta, and M. Sanjappa, "Plant wealth of northeast india with reference to ethnobotany," 2009, https://www.researchgate.net/publication/283868339_Plant_wealth_of_Northeast_India_with_reference_to_ethnobotany.
- [28] N. Borah, F. D. Athokpam, R. Semwal, and S. Garkoti, "(Black Rice; *Oryza sativa* L.): a culturally important and stress tolerant traditional rice variety of Manipur," 2018, https://www.academia.edu/40446536/Chakhao_Black_Rice_Oryza_sativa_L._A_culturally_important_and_stress_tolerant_traditional_rice_variety_of_Manipur.
- [29] I. D. Asem, A. Nongthombam, K. Shaheen et al., "Phenotypic characterization, genetic variability and correlation studies among ten Chakhao (scented) rice of Manipur," *International Journal of Current Microbiology and Applied Sciences*, vol. 8, no. 2, pp. 612–618, 2019.
- [30] S. Bhuvanawari, S. Gopala Krishnan, R. K. Ellur et al., "Discovery of a novel induced polymorphism in SD1 gene governing semi-dwarfism in rice and development of a functional marker for marker-assisted selection," *Plants*, vol. 9, 2020.
- [31] V. R. Hinge, H. B. Patil, and A. B. Nadaf, "Aroma volatile analyses and 2AP characterization at various developmental stages in Basmati and Non-Basmati scented rice (*Oryza sativa* L.) cultivars," *Rice*, vol. 9, no. 1, pp. 38–22, 2016.
- [32] R. Bryant and A. McClung, "Volatile profiles of aromatic and non-aromatic rice cultivars using SPME/GC-MS," *Food Chemistry*, vol. 124, no. 2, pp. 501–513, 2011.
- [33] S. Cho, E. Nuijten, R. L. Shewfelt, and S. J. Kays, "Aroma chemistry of African *oryza glaberrima* and *oryza sativa* rice and their interspecific hybrids," *Journal of the Science of Food and Agriculture*, vol. 94, no. 4, pp. 727–735, 2014.
- [34] M. H. Givianrad, "Characterization and assessment of flavor compounds and some allergens in three Iranian rice cultivars during gelatinization process by HS-SPME/GC-MS," *E-Journal of Chemistry*, vol. 9, no. 2, pp. 716–728, 2012.
- [35] A. Griglione, E. Liberto, C. Cordero et al., "High-quality Italian rice cultivars: chemical indices of ageing and aroma quality," *Food Chemistry*, vol. 172, pp. 305–313, 2015.
- [36] K. Mahattanatawee and R. L. Rouseff, "Comparison of aroma active and sulfur volatiles in three fragrant rice cultivars using GC-Olfactometry and GC-PFPD," *Food Chemistry*, vol. 154, pp. 1–6, 2014.
- [37] C. Grimm and E. T. Champagne, "Analysis of volatile compounds in the headspace of rice using SPME/GC/MS," *Flavor, fragrance and odor analysis*, pp. 229–248, 2001.
- [38] J.-Y. Lin, W. Fan, Y.-N. Gao, S.-F. Wu, and S.-X. Wang, "Study on volatile compounds in rice by HS-SPME and GC-MS," *Julius-Kühn-Archiv*, vol. 425, p. 125, 2010.
- [39] D. Weber, R. Rohilla, and U. Singh, *Chemistry and Biochemistry of Aroma in Scented Rice*, pp. 29–46, Oxford & IBH Publ, New Delhi, India, 2000.
- [40] M. Calingacion, L. Fang, L. Quiatchon-Baeza et al., "Delving deeper into technological innovations to understand differences in rice quality," *Rice*, vol. 8, no. 1, pp. 6–10, 2015.
- [41] T. Piyachaiseth, W. Jirapakkul, and S. Chaiseri, "Aroma compounds of flash-fried rice," *Agriculture and Natural Resources*, vol. 45, no. 4, pp. 717–729, 2011.
- [42] W. Setyaningsih, T. Majchrzak, T. Dymerski, J. Namieśnik, and M. Palma, "Key-marker volatile compounds in aromatic rice (*Oryza Sativa*) Grains: an HS-SPME extraction method combined with GC×GC-TOFMS," *Molecules*, vol. 24, no. 22, p. 4180, 2019.
- [43] S. Mahatheeranont, S. Keawsa-ard, and K. Dumri, "Quantification of the rice aroma compound, 2-acetyl-1-pyrroline, in uncooked Khao Dawk Mali 105 brown rice," *Journal of Agricultural and Food Chemistry*, vol. 49, no. 2, pp. 773–779, 2001.
- [44] U. Attar, V. Hinge, R. Zanan, R. Adhav, and A. Nadaf, "Identification of aroma volatiles and understanding 2-acetyl-1-pyrroline biosynthetic mechanism in aromatic mung bean (*Vigna radiata* (L.) Wilczek)," *Physiology and Molecular Biology of Plants*, vol. 23, no. 2, pp. 443–451, 2017.
- [45] D. K. Verma and P. P. Srivastav, "Extraction, identification and quantification methods of rice aroma compounds with emphasis on 2-acetyl-1-pyrroline (2-AP) and its relationship with rice quality: a comprehensive review," *Food Reviews International*, vol. 38, no. 2, pp. 111–162, 2020.
- [46] C. C. Lswg, "Flavor profiles of aromatic and non-aromatic rice varieties," in *Proceedings of the PITTCON Conference and ExpoPaper presented at: Pittsburgh Conference, Orlando, FL, USA, 2010*.
- [47] P. Bhattacharjee, A. Kshirsagar, and R. S. Singhal, "Super-critical carbon dioxide extraction of 2-acetyl-1-pyrroline from

- Pandanus amaryllifolius Roxb,” *Food Chemistry*, vol. 91, no. 2, pp. 255–259, 2005.
- [48] M. L. Wu, K. L. Chou, C. R. Wu, J. K. Chen, and T. C. Huang, “Characterization and the possible formation mechanism of 2-acetyl-1-pyrroline in aromatic vegetable soybean (*Glycine max* L.),” *Journal of Food Science*, vol. 74, no. 5, pp. S192–S197, 2009.
- [49] M. A. Majcher, D. Klensporf-Pawlik, M. Dziadas, and H. H. Jeleń, “Identification of aroma active compounds of cereal coffee brew and its roasted ingredients,” *Journal of Agricultural and Food Chemistry*, vol. 61, no. 11, pp. 2648–2654, 2013.
- [50] M. Calingacion, A. Laborte, A. Nelson et al., “Diversity of global rice markets and the science required for consumer-targeted rice breeding,” *PLoS One*, vol. 9, no. 1, Article ID e85106, 2014.
- [51] C. S. Chanu, N. Yenagi, and K. Math, “Nutritional and functional evaluation of black rice genotypes,” *Journal of Farm Sciences*, vol. 29, no. 1, pp. 61–64, 2016.
- [52] Q. Xie, F. Li, L. Fang, W. Liu, and C. Gu, “The antitumor efficacy of β -elemene by changing tumor inflammatory environment and tumor microenvironment,” *BioMed Research International*, vol. 2020, Article ID 6892961, 13 pages, 2020.
- [53] J.-J. Lu, Y.-Y. Dang, M. Huang, W.-S. Xu, X.-P. Chen, and Y.-T. Wang, “Anti-cancer properties of terpenoids isolated from *Rhizoma Curcumae*—a review,” *Journal of Ethnopharmacology*, vol. 143, no. 2, pp. 406–411, 2012.